



DEVELOPMENTAL STUDIES HYBRIDOMA BANK

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F5D

INVESTIGATOR

Name Woodring E. Wright, Ph.D.

Address University of Texas Southwestern Medical Center, 5323 Harry Hines Blvd., Dallas, TX 75235-9039

IMMUNOGEN

Substance

Name myogenin

Origin rat

Chemical Composition glutathione-s-transferase fusion protein

Developmental Stage no known isoforms

IMMUNIZATION PROTOCOL

Donor Animal

Species mouse

Strain BALB/c

Sex

Organ and tissue popliteal lymphocytes

Immunization

Dates immunized May 19 & 22 (RIBI), May 26, 30, June 2 & 5 (PBS) 1989

Amount of antigen 50µg Gst-myogenin per site

Route of immunization rear foot pads

Adjuvant RIBI for 1st two injections, then in PBS

FUSION

Date

Myeloma cell line

Species mouse

Designation NS1

MONOCLONAL ANTIBODY

Isotype IgG1, kappa light chain

Specificity

Cell binding no (antigen is internal)

Immunohistology yes

Antibody competition untested

Species Specificity mammalian (tested on rat, mouse, cat, human; quail did not react)

ANTIGEN

Chemical properties

rat myogenin

Molecular weight

~ 25 kDa; migrates as ~ 34 kDa on SDS gels; 224 a.a.

Characterization

Immunoprecipitation immunoprecipitates deletion mutants containing a.a. 138-158, a region immediately carboxy-terminal to the bHLH domain.

Immunoblotting positive

Purification cloned

Amino acid sequence analysis deduced from cDNA

Functional effects skeletal muscle-specific bHLH transcription factor probably controlling myoblast to myotube transition.

Immunohistochemistry nuclear

PUBLICATIONS :

Wright, W.E., Bender, M., Funk, W. (1991). Cyclic amplification and selection of targets (CASTing) for the myogenin consensus binding site. Mol. Cell. Biol. 11, 4104-4110.

Cusella-De Angeles, M.G., Lyons, G., Sonnino, C., De Angelis, L., Vivarelli, E., Wright, W.E., Molinaro, M., Bouché, M., Buckingham, M. and Cossu, G. (1992). MyoD1, myogenin independent differentiation of primordial myoblasts in somites. J. Cell Biol. 116, 1243-1255.

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**F5D** (Continued)

- Yablonka-Reuveni, Z., and Rivera, A.J. (1994). Temporal expression of regulatory and structural muscle proteins during myogenesis of satellite cells on isolated adult rat fibers. *Dev. Biol.* 164, 588-603.
- Wang, N.P., Marx, J., McNutt, M.A., Rutledge, J.C., and Gown, A.M. (1995). Expression of myogenic regulatory proteins (myogenin and myoD1) in small blue round cell tumors of childhood. *Am. J. of Path.* 147, 1799-1810.
- Yablonka-Reuveni, Z. (1995). Development and postnatal regulation of adult myoblasts. *Microsc. Res. Tech.* 30, 366-380.
- Wright, W.E., Dac-Korytko, I., and Farmer, K. (1996). Monoclonal antimyogenin antibodies define epitopes outside the bHLH domain where binding interferes with protein-protein and protein-DNA interactions. *Dev. Genetics* 19, 131-138.
- Yablonka-Reuveni, Z., and Rivera, A.J. (1997). Influence of PDGF-BB on proliferation and transition through the MyoD-myogenin-MEF2A expression program during myogenesis in mouse C2 myoblasts. *Growth Factors* 15, 1-27.
- Dominov, J.A., Dunn, J.J., and Miller, J.B. (1998). Bcl-2 expression identifies an early stage of myogenesis and promotes clonal expansion of muscle cells. *J. Cell Biol.* 142, 537-544.
- Naro, F., Sette, C., Vicini, E., De Arcangelis, V., Grange, M., Conti, M., Lagarde, M., Molinaro, M., Adamo, S., and Nemoz, G. (1999). Involvement of type 4 cAMP-phosphodiesterase in the myogenic differentiation of L6 cells. *Mol. Biol. Cell* 10, 4355-4367.
- Dube, M., Huot, M.-E., and Khandjian, E.W. (2000). Muscle specific fragile X related protein I isoforms are sequestered in the nucleus of undifferentiated myoblast. *BMC Genetics*, 1(4). Retrieved May 20, 2001 on the World Wide Web: <http://www.biomedcentral.com/1471-2156/1/4>
- Young, H.E., Duplaa, C., Young T.M., Floyd, J.A., Reeves, M.L., Davis, K.H., Mancini, G.J., Eaton, M.E., Hill, J.D., Thomas, K., Austin, T., Edwards, C., Cuzzourt, J., Parikh, A., Groom, J., Hudson J., and Black, Jr., A.C. (2001). Clonogenic analysis reveals reserve stem cells in postnatal mammals: I. Pluripotent mesenchymal stem cells. *Anat. Rec.* 263, 350-360.
- Young, H.E., Steele, T.A., Bray, R.A., Hudson, J., Floyd, J.A., Hawkins, K., Thomas, K., Austin, T., Edwards, C., Cuzzourt, J., Duenzl, M., Lucas, P.A., and Black, Jr., A.C. (2001). Human reserve pluripotent mesenchymal stem cells are present in the connective tissues of skeletal muscle and dermis derived from fetal, adult, and geriatric donors. *Anat. Rec.* 264, 51-62.
- Hirayama, E., Udaka, Y., Kawai, T., and Kim, J. (2001). Characterization of heterokaryons between skeletal myoblasts and somatic cells formed by fusion with HVJ (Sendai virus); effects on myogenic differentiation. *Cell Struct. Funct.* 26, 37-47.
- Dedkov, E.I., Kostrominova, T.Y., Borisov, A.B., and Carlson, B.M. (2001). Reparative myogenesis in long-term denervated skeletal muscles of adult rats results in a reduction of the satellite cell population. *Anat. Rec.* 263, 139-154.
- Sasao, N., Hirayama, E., and Kim, J. (2003). Characterization of heterokaryons between skeletal myoblasts and preadipocytes: myogenic potential of 3T3-L1 preadipocytes. *Eur. J. Cell Biol.* 82, 97-103.
- Dedkov, E.I., Kostrominova, T.Y., Borisov, A.B., and Carlson, B.M. (2003). MyoD and myogenin protein expression in skeletal muscles of senile rats. *Cell Tissue Res.* 311, 401-416.
- De Arcangelis, V., Coletti, D., Conti, M., Lagarde, M., Molinaro, M., Adamo, S., Nemoz, G., and Naro, F. (2003). IGF-I-induced differentiation of L6 myogenic cells requires the activity of cAMP-phosphodiesterase. *Mol. Biol. Cell* 14, 1392-1404.
- Naro, F., De Arcangelis, V., Sette, C., Ambrosio, C., Komati, H., Molinaro, M., Adamo, S., and Nemoz, G. (2003). A bimodal modulation of the cAMP pathway is involved in the control of myogenic differentiation in L6 cells. *J. Biol. Chem.* 278(49), 49308-49315.
- Komati, H., Minasi, A., Naro, F., Lagarde, M., Prigent, A.-F., Adamo, S., and Nemoz, G. (2004). Phorbol ester-induced differentiation of L6 myogenic cells involves phospholipase D activation. *FEBS Lett.* 577, 409-414.
- Young, H.E. (2004). Existence of reserve quiescent stem cells in adults, from amphibians to humans. *Curr. Top. Microbiol. Immunol.* 280, 71-109.
- Young, H.E., and Black, Jr., A.C. (2004). Adult stem cells. *Anat. Rec. Part A* 276A, 75-102.
- Mendias, C.L., Tatsumi, R., and Allen, R.E. (2004). Role of cyclooxygenase-1 and -2 in satellite cell proliferation, differentiation, and fusion. *Muscle Nerve* 30, 497-500.
- Shefer, G., Wleklinski-Lee, M., and Yablonka-Reuveni, Z. (2004). Skeletal muscle satellite cells can spontaneously enter an alternative mesenchymal pathway. *J. Cell Sci.* 117, 5393-5404.
- Komati, H., Naro, F., Mebarek, S., De Arcangelis, V., Adamo, S., Lagarde, M., Prigent, A.-F., and Nemoz, G. (2005). Phospholipase D is involved in myogenic differentiation through remodeling of actin cytoskeleton. *Mol. Biol. Cell* 16, 1232-1244.
- Schuster-Gossler, K., Cordes, R., and Gossler, A. (2007). Premature myogenic differentiation and depletion of progenitor cells cause severe muscle hypotrophy in Delta1 mutants. *PNAS* 104(2), 537-542.
- Coletti, D., Teodori, L., Albertini, M.C., Rocchi, M., Pristera, A., Fini, M., Molinaro, M., and Adamo, S. (2007). Static magnetic fields enhance skeletal muscle differentiation in vitro by improving myoblast alignment. *Cytometry A* 71A, 846-856.
- Makarenkova, H.P., Gonzalez, K.N., Kiosses, W.B., and Meech, R. (2009). Barx2 controls myoblast fusion and promotes MyoD-mediated activation of the smooth muscle α -actin gene. *J. Biol. Chem.* 284(22), 14866-14874.

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F5D (Continued)

- Otto, A., Macharia, R., Matsakas, A., Valasek, P., Mankoo, B.S., and Patel, K. (2010). A hypoplastic model of skeletal muscle development displaying reduced foetal myoblast cell numbers, increased oxidative myofibres and improved specific tension capacity. *Dev. Biol.* 343, 51-62.
- Voronova, A., Coyne, E., Al Madhoun, A., Fair, J.V., Bosiljcic, N., St-Louis, C., Li, G., Thurig, S., Wallace, V.A., Wiper-Bergeron, N., and Skerjanc, I.S. (2013). Hedgehog signaling regulates MyoD expression and activity. *J. Biol. Chem.* 288(6), 4389-4404. doi:10.1074/jbc.M112.400184.
- Garcia-Parra, P., Naldaiz-Gastesi, N., Maroto, M., Padin, J.F., Goicoechea, M., Aiastui, A., Fernandez-Morales, J.C., Garcia-Belda, P., Lacalle, J., Alava, J.I., Garcia-Verdugo, J.M., Garcia, A.G., Izeta, A., and Lopez de Munain, A. (2013). Murine muscle engineered from dermal precursors: an in vitro model for skeletal muscle generation, degeneration, and fatty infiltration. *Tiss. Eng.* 20(1), doi:10.1089/ten.tec.2013.0146.
- Ciarapica, R., De Salvo, M., Carcarino, E., Bracaglia, G., Adesso, L., Leoncini, P.P., Dall'Agnese, A., Walters, Z.S., Verginelli, F., De Sio, L., Boldrini, R., Inserra, A., Bisogno, G., Rosolen, A., Alaggio, R., Ferrari, A., Collini, P., Locatelli, M., Stifani, S., Screpanti, I., Rutella, S., Yu, Q., Marquez, V.E., Shipley, J., Valente, S., Mai, A., Miele, L., Puri, P.L., Locatelli, F., Palacios, D., and Rota, R. (2013). The polycomb group (PcG) protein EZH2 supports the survival of PAX3-FOXO1 alveolar rhabdomyosarcoma by repressing FBXO32(Atrogin1/MAFbx). *Oncogene* doi: 10.1038/onc.2013.471.

ACKNOWLEDGMENTS STATEMENT

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F5D

Anti-rat Myogenin, monoclonal antibody, lab name originally 1F5D7, renamed F5D for short.

Heavy chain: IgG1 Light chain: K

Epitope: between aa 144-170 (Helix 2 ends at aa 132, the last of the three leucines in the potential leucine zipper is aa 146)

Our growth conditions: RPMI1640 + 10% FCS + 25 µg/ml gentamicin

Our harvest conditions: Seed cells at 1 million/ml in either HB101 (for serum free) or 2% FCS: Harvest medium q24 hours for 2-3 days until cells have died, counting and reseeded at 1 million/ml each time.

To Freeze: (if cells sent): resuspend cells at one million/ml in 90% serum/10% DMSO. Put inside a cardboard box (like a cell inventory freezer box, approx. 12 cm x 12 cm x 5 cm) and put directly at -80°C overnight or longer, then transfer to -135°C or below.

Known efficacy:

Immunofluorescence: Cells fixed 20 min at room temp in 2% paraformaldehyde in PBS, then permeabilized 10 min at room temp with 0.15% triton X-100 in PBS. We've only done it with neat culture supernatant, but it probably would work if diluted some.

Western Blotting: Good signal with 1:10 diluted culture supernatant.

Gel Shift: Gives good double-shift of myogenin in nuclear extracts using 0.1 µl of a 50-fold concentrated supernatant (i.e., the equivalent of 5 µl of supernatant. Less might still work, but that was the lowest we tried).

Conditions for acceptance of this reagent:

- 1) Use them for anything you want, but please let me know anything interesting that you find. If you are using them for something other than as a marker etc. that you think is likely to be in direct competition with what I'm doing, it would be nice (but not required) to let me know and we'll discuss whether or not it should be collaborative etc.
- 2) If you make a derivative reagent (i.e., you purify the antibody and directly label it with anything [fluorescein, rhodamine, HRP, gold, etc]), let me know and agree to provide me with an aliquot if it would be useful to me.
- 3) You may provide the cells/supernatants to third parties as long as you provide a copy of this letter giving the conditions of use.

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